

Molecular Docking Studies on the Interaction of Anti-Alzheimer Compounds with Amyloid Beta Peptides

Nur Hana Faujan¹

¹Center of Foundation Studies for Agricultural Science, Universiti Putra Malaysia, 43400 Serdang UPM, Selangor, Malaysia
Email address: nurhana@upm.edu.my

Norzalina Zakaria² and Nurul Najihah Mohammad²

²Department of Chemistry, Faculty of Science, Universiti Putra Malaysia, 43400 UPM, Serdang, Selangor, Malaysia

Abstract—Alzheimer's disease (AD) is an irreversible, progressive neurodegenerative disorder that slowly affected to the loss of memory and the ability to perform simple cognitive activities. Amyloid Beta (A β) peptide catalysed by β - and γ -secretases is the main hypothesis of AD progression that is associated with synaptic failure in brain. The A β_{1-42} is the most prevalent form of A β peptides that have been suggested as a primary key in AD. Molecular docking studies have been performed to investigate the interaction of A β with nine molecules, which were found to exhibit amyloid inhibitory effect. We found that folic acid and donepezil compounds showed high binding affinity in both A β_{42} peptide (PDB ID: 1IYT and 1Z0Q), in comparison to the other ligands. The binding interactions of folic acid and donepezil compounds with active site of A β protein model suggested that the amino acid residues of Glu11, Phe19, Gln15, Glu22 and Val12 play a key role for drug design.

Keywords—molecular docking, amyloid β peptide, hydrogen bonding, Alzheimer's

Introduction

Alzheimer's disease (AD) is a neurological disorder characterised by the aggregation of amyloid beta (A β) protein into fibrillary amyloid plaques in the brain. The A β peptide is the product of amyloid precursor protein (APP) proteolysis, mostly composed of 42 amino acids (A β_{1-42}) sequence [1]. The A β_{42} produced small neurotoxic oligomers, which is the main pathogenic peptide in AD. Thus, the inhibition of A β peptide is one of the strategy in designing potential drug for AD therapy.

This study was aimed to analyze the interactions of two forms of A β_{42} peptide with nine ligands. Five molecules were selected based on their reported inhibitory effect on A β either *in vitro* or *in vivo* which are good candidates for AD treatment. These compounds are curcumin, folic acid, ibuprofen, rosmarinic acid and ascorbic acid. The rest were drugs approved by the U.S. Food and Drug Administration (FDA) to treat Alzheimer's patients. Current drugs for AD such as donepezil, galantamine, rivastigmine and

tacrine are acetylcholinesterase inhibitors that can treat some symptoms but not to cure the disease.

Ibuprofen as a non-steroidal anti-inflammatory drugs has been found to control the A β aggregation [Zurita, 2013]. Natural compounds such as curcumin, folic acid, rosmarinic acid and ascorbic acid are nontoxic with high anti-oxidant properties [Rebekah, 2015]. Both curcumin and rosmarinic acid compounds have been found to inhibit the formation of amyloid fibril *in vitro* [2]. Recently, folic acid has been examined to improve cognition and inflammation markers on AD patients [Chen, 2016]. Other study reported that ascorbic acid was found to reduce amyloid plaque burden *in vivo* [Kook, 2014].

Although some compounds including curcumin and folic acid are under clinical trial phase, their binding interactions with A β peptides have not yet been understood at the molecular level. Computer simulation such as molecular docking has been successfully used for studying protein-ligand interaction and virtual screening. Molecular docking was performed between aminopeptidase (SGAK) with 1IYT peptide showed that Glu139 might play an important role to degrade A β peptide [Dhanavade, 2014]. AutoDock Vina was applied to investigate the binding of ergothionein and selenoergothionein as a potential inhibitor against 1IYT peptide [Saddala, 2016]. The interactions of anti-amyloidogenic behaviour of 40 small molecule inhibitors with A β_{1-40} and Iowa mutant D₂₃N-A β_{15-42} peptides and their modes of binding have been studied using molecular docking approaches [Khan, 2019].

Docking results are very useful to predict the behaviour of the compounds in the binding site of targets as well as to explain fundamental biochemical processes [3]. Here, AutoDock Vina was used for studying the binding interaction of nine compounds including curcumin, donepezil, folic acid, galantamine, ibuprofen, rosmarinic acid, rivastigmine, tacrine and ascorbic acid. The docking complexes were analysed based on their binding pose, binding affinity and interaction.

Methodology

Protein preparation

The three-dimensional (3D) structure of the A β_{42} peptide in a polar environment (PDB ID: 1IYT)

(Crescenzi, 2002) and apolar environment (PDB ID: 1Z0Q) (Tomaselli, 2006) were taken from the Protein Data Bank (PDB) (<http://www.rcsb.org/pdb>).

Ligand preparation

The structures of all the compounds in TABLE I were obtained from PubChem database [4] in SDF format and converted into PDB using Discovery Studio Visualization [5] for further docking studies. The structures were minimised using general AMBER force field (GAFF) in Avogadro software [6].

TABLE I. THE MOLECULAR STRUCTURE AND MOLECULAR WEIGHT OF LIGANDS

Ligand Compounds	PubChem ID	Molecular Formula	Molecular weight (g/mol)
Curcumin	969516	C ₂₁ H ₂₀ O ₆	368.40
Donepezil	3152	C ₂₄ H ₂₉ NO ₃	379.50
Folic Acid	135398658	C ₁₉ H ₁₉ N ₇ O ₆	441.40
Galantamine	9651	C ₁₇ H ₂₁ NO ₃	287.35
Ibuprofen	3672	C ₁₃ H ₁₈ O ₂	206.28
Rosmarinic acid	5281791	C ₁₈ H ₁₆ O ₈	360.31
Rivastigmine	77991	C ₁₄ H ₂₂ N ₂ O ₂	250.34
Tacrine	1935	C ₁₃ H ₁₄ N ₂	198.26
Ascorbic acid	54670067	C ₆ H ₈ O ₆	176.12

Molecular docking

Docking studies yielded crucial information concerning the orientation of ligand in the binding pocket of the target protein. Each compound was docked against the structure of A β ₄₂ monomer in both polar (hereafter referred as 1IYT receptor) and apolar (hereafter referred to as 1Z0Q receptor) to determine their binding affinity using AutoDock Vina [Trott, Haas, 2007]. The grid and docking parameter files were prepared using AutoDock Tools (ADT) [7]. The grid boxes with dimensions of 126 x 74 x 126 Å with grid spacing 0.403 Å and 126 x 82 x 76 Å with grid spacing 0.419 Å were defined for 1IYT and 1Z0Q respectively. These customized dimensions are large enough to cover the whole protein and leave enough space for ligands to be docked on surface.

The two-dimensional graphical depictions of best docked complexes were accessed using Ligplot+v.1.4.5 [Wallace,1995, Laskowski,2011, 8] and detailed analysis on the physicochemical of the docked complexes were obtained from PLIP: fully automated protein–ligand interaction profiler [9].

Results and Discussion

Molecular docking has been used to predict most stable conformation of a non-covalently bound molecule towards the other one as well as its binding affinity. Binding affinity is the strength of the binding interaction between a single biomolecule to its ligand, which used to estimate and rank order strengths of biomolecular interactions. It is influenced by non-covalent intermolecular interaction such as hydrogen bonding, hydrophobic, electrostatic interactions and van der Waals forces between the two molecules.

In this study, AutoDock Vina was utilised to estimate the mode corresponds to the binding affinity between the ligands and proteins. The binding affinity predictability of AutoDock Vina was correlated with the experimental binding affinity corresponded with non-specific van der Waals and hydrophobic interaction scores between ligands and receptors [10].

The binding affinity obtained in the best mode are shown in TABLE II. Folic acid (MW=441.40 g/mole) and Donepezil (MW=379.49 g/mole) show high binding affinity to A β ₄₂ monomer in both polar and apolar environment, in comparison to the other ligands. The weakest binding affinity was observed for ascorbic acid (MW=176.12 g/mole). The protein-ligand binding energy tend to correlate with the molecular weight of the ligands to some extent.

TABLE II. BINDING AFFINITY OF ANTI-ALZHEIMER COMPOUNDS WITH 1IYT AND 1Z0Q RECEPTORS

Ligand name	Targeted A β ₄₂ Peptides	Binding Affinity (kcal/mol)
Curcumin	1IYT	-4.5
	1Z0Q	-5.1
Donepezil	1IYT	-5.2
	1Z0Q	-6.3
Folic Acid	1IYT	-5.6
	1Z0Q	-6.0
Galantamine	1IYT	-5.4
	1Z0Q	-5.3
Ibuprofen	1IYT	-4.7
	1Z0Q	-4.5
Rosmarinic acid	1IYT	-4.3
	1Z0Q	-5.5
Rivastigmine	1IYT	-4.0
	1Z0Q	-4.3
Tacrine	1IYT	-5.1
	1Z0Q	-5.7
Ascorbic acid	1IYT	-3.8
	1Z0Q	-3.9

Molecular docking studies were done to investigate the interaction of anti-Alzheimer's molecules with A β ₄₂. TABLE III shows the hydrogen bonding, hydrophobic and π -stacking interactions of the docked complexes. Folic acid forms favorable hydrogen bonding and hydrophobic interaction with both 1IYT and 1Z0Q receptors. Surprisingly, the hydrogen bond, hydrophobic and π -stacking interactions exist between folic acid compound and A β ₄₂ peptide in apolar environment. Three types of interactions such as hydrogen bonds, hydrophobic and π -stacking exist between donepezil and A β ₄₂ receptors.

TABLE III. THE NUMBER OF HYDROGEN BONDING, HYDROPHIC AND π -STACKING FORMED IN PROTEIN-LIGANDS INTERACTIONS

Ligand name	Targeted A β ₄₂ Peptide	H-bond	Hydrophobic	π -Stacking
Curcumin	1IYT	2	4	1
	1Z0Q	3	4	1
Donepezil	1IYT	1	3	1
	1Z0Q	1	2	1
Folic Acid	1IYT	5	3	1
	1Z0Q	8	2	0

Galantamine	1IYT	1	3	0
	1Z0Q	3	2	0
Ibuprofen	1IYT	1	3	1
	1Z0Q	0	4	0
Rosmarinic acid	1IYT	1	3	1
	1Z0Q	1	2	1
Rivastigmine	1IYT	0	3	1
	1Z0Q	1	2	1
Tacrine	1IYT	2	3	1
	1Z0Q	2	4	0
Ascorbic acid	1IYT	3	0	0
	1Z0Q	5	0	0

Hydrogen bond interactions are crucial to ligand protein binding. Folic acid showed the highest interaction by forming two hydrogen bonds with Glu3 and Gly9, and one hydrogen bond between His13 residues of 1IYT receptor. In 1Z0Q receptor, it forms eight hydrogen bonds; two hydrogen bonds with Glu11, Gln15 and Asn27, and one hydrogen bond with Glu22, and Asp23. A hydrogen bond exists against donepezil with Gln15 residue in both 1IYT and 1Z0Q receptors.

As shown in Fig. 1, one hydrogen bond was observed between the NH group on the quinazoline ring, and the other NH group of the folic acid with the carboxyl groups of Glu3 at distance of 2.97 Å and 3.69 Å, respectively. The third and fourth hydrogen bonds were observed between NH and carboxyl groups of folic acid with the carboxyl groups of Gly9 at distance of 3.17 Å and 2.99 Å, respectively. In addition, folic acid forms one hydrogen bond between NH with the carboxyl group of His13 at distance of 3.89 Å.

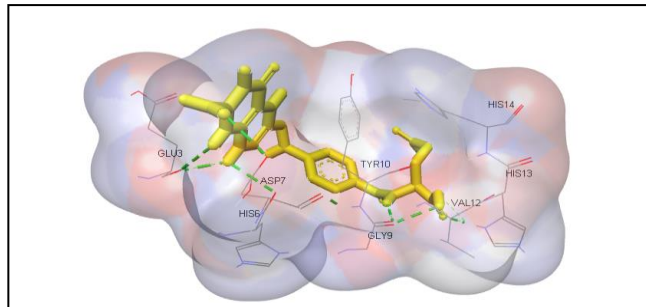


Fig. 1. 3D structure of folic acid docked to 1IYT receptor

Donepezil interact via hydrogen bond through its NH of amine group with Gln15 at distance 3.76 Å (Fig. 2).

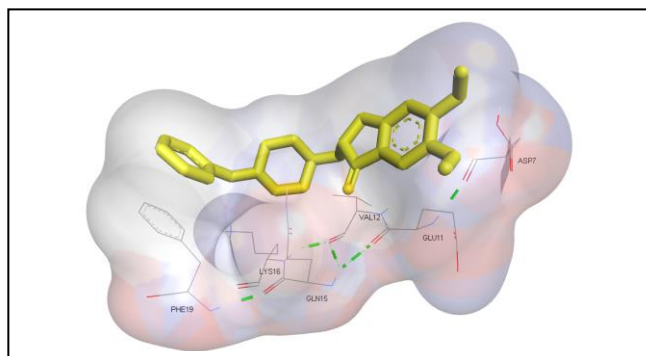


Fig. 2. 3D structure of donepezil docked to 1IYT receptor

Folic acid has some hydrophobic interaction with two amino acids of His6 and Thr10 against 1IYT receptor; the His6 forms one hydrophobic interaction at distance 3.85 Å and Thr10 forms two hydrophobic interactions at distance 3.68 Å and 3.66 Å, respectively. In addition, folic acid interacts via π -stacking bonds with Try10.

Donepezil interacts via π -stacking bond with Phe19 and some hydrophobic interactions; the Glu11 forms one hydrophobic interaction at distance 3.74 Å and Val12 forms two hydrophobic interactions at distance 3.93 Å and 3.46 Å, respectively (TABLE III).

In 1Z0Q receptor, folic acid interact via hydrogen bonds through its NH group on the quinazoline ring and the other NH group of the folic acid with carboxyl groups of Glu11 at distance 3.17 Å and 2.86 Å, respectively. The two hydrogen bonds exist between NH of folic acid moiety with the NH₂ groups of Gln15 at distance 3.76 Å and 3.64 Å, respectively. The other NH and carbonyl groups of folic acid form hydrogen bonds between the carbonyl groups of Asn27 at distance 3.02 Å and 3.79 Å, respectively. In 1Z0Q docked complex, carboxyl group of folic acid interact by hydrogen bond with Glu22, and Asp23 at a distance of 3.05 Å and 3.06 Å, respectively (Fig. 3).

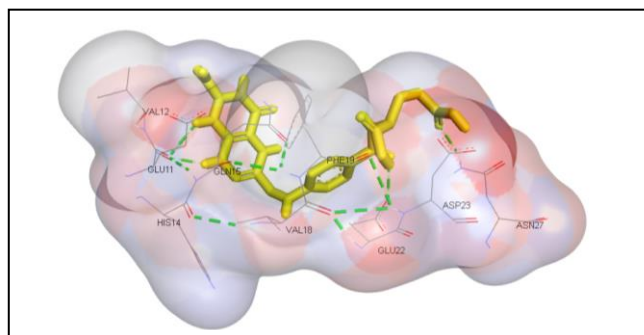


Fig. 3. 3D structure of folic acid docked to 1IYT receptor

As shown in Fig. 3, one hydrogen bond was observed between the NH amine of donepezil with Gln15 at distance of 3.10 Å in 1I20Q docked complex.

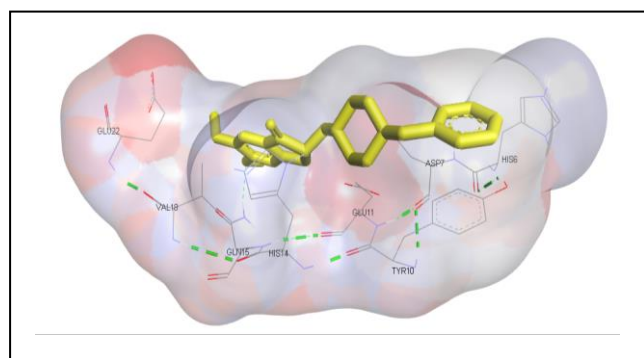


Fig. 4. 3D structure of donepezil docked to 1Z0Q receptor

In 1Z0Q receptor, folic acid has two hydrophobic interactions with two hydrophobic amino acids of Val18 and Glu22 at distance 3.98 Å and 3.68 Å, respectively. Donepezil interacts via two hydrophobic interactions with Glu11 and Val18 at distance 3.79 Å and 3.65 Å, respectively. There is also a π -stacking bond with Tyr10 residue of 1Z0Q receptor.

Folic acid is one of the B-complex vitamins with an aromatic compounds contains two carboxyl groups and one aryl ring join to a dihydropteridine ring. Based on our result, it's highly interacting with Glu11, Glu22, Phe19 and Asp23 in both 1IYT and 1Z0Q receptors. This suggests a possible amyloid inhibitory mechanism of folic acid compound by binding and masking amyloidogenic region in the peptide, thus preventing self-aggregation of A β peptide. Furthermore, the essential role of hydrogen bonding and hydrophobic interaction assist the binding of folic acid towards A β peptide, which might prevent further elongation of the A β fibrils.

Conclusion

Alzheimer is the most common forms of dementia characterized by deposition of A β peptide. Several natural compounds have been used to improve memory and cognition in AD patients as supported by various experimental studies. Docking study that was applied to analyse the best docked ligands permitted us to know the binding mode of compounds to the target Alzheimer peptides. The amino acids that contribute the most to the interaction of folic acid and donepezil compounds with active site of A β protein model are Glu11 and Phe19, followed by Gln15, Glu 22 and Val12. This study found that folic acid has strong interactions with A β peptide by forming more hydrogen bonds, hydrophobic as well as π -stacking interactions with both 1IYT and 1Z0Q receptors. The findings from this study might assist researchers in designing A β inhibitors and formulate folic acid supplement as a treatment for AD.

ACKNOWLEDGMENT

The authors gratefully acknowledge financial support from GP-IPM Vote Number: 9628900 grant from Universiti Putra Malaysia.

REFERENCES

- [1] Hardy, J. and Selkoe. D.J., 2002. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*, 297:5580, 353-356.
- [2] Zurita, M.P., Munoz, G., Sepulveda F.J., Gome, P., Castillo, C., Fuentealba J., Opazo, C., Aguayo, L.G. 2013. Ibuprofen inhibits the synaptic failure induced by the amyloid- β peptide in hippocampal neurons. *Journal of Alzheimer's Disease*, 35(3),463-473.
- [3] Watkins, R., Wu, L., Zhang, C., Davis, R.M., and Xu, B., 2015. Natural product-based nanomedicine: recent advances and issues. *International Journal of Nanomedicine* 10, 655-6074.
- [4] Porat, Y., Abramowitz, A. and Gazit. E., 2006. Inhibition of amyloid fibril formation by polyphenols: structural similarity and aromatic interactions as common inhibition mechanism. *Chem Biol Drug Des.* 67:27-37.
- [5] Chen H., Liu S., Ji L., Tianfeng W., Ji Y., Zhou Y., Zheng M., Zhang M., Xu M., and Huang G., 2016. Folic Acid Supplementation Mitigates Alzheimer's

Disease by Reducing Inflammation: A Randomized Controlled Trial. *Mediators of Inflammation*. Article ID 5912146.

[6] Marcus D Hanwell, Donald E Curtis, David C Lonie, Tim Vandermeersch, Eva Zurek and Geoffrey R Hutchison, 2012. Avogadro: An advanced semantic chemical editor, visualization, and analysis platform. *Journal of Cheminformatics*, 4:17.

[7] Dhanavade, M.J and Sonawane, K.D. 2014. Insights into the molecular interactions between aminopeptidase and amyloid beta peptide using molecular modeling techniques. *Amino Acids*. 46(8):1853-66.

[8] Saddala, M.S., Adi, P. K. J., Patchava, V. R., and Usha Rani, A., 2016. Selenoergothionein as a Potential Inhibitor against Amyloid β -Protein (A β): Docking and Molecular Dynamics Studies. *Int J Biomed Data Min*, 5:2.

[9] Khan, R.H, Siddqi, M.K., Uversky, V.N and Salahuddin, P., 2019. Molecular docking of A β ₁₋₄₀ peptide and its Iowa D₂₃N mutant using small molecule inhibitors: Possible mechanisms of A β -peptide inhibition. *Int J Biol Macromol.* 15;127:250-270.

[10] Meng, X-Y, Zhang, H-X, Mezei, M., and Cui, M., 2011. Molecular Docking: A Powerful Approach for Structure-Based Drug Discovery, *Curr. Comput. Aided-Drug Des.*, 7:2, 146-157.

[11] Crescenzi O., Tomaselli S., Guerrini R., Salvadori S., D'Ursi A.M., Temussi P.A, Picone, D., 2002. Solution Structure of the Alzheimer Amyloid Beta-peptide (1-42) in an Apolar Microenvironment. Similarity with a Virus Fusion Domain, *European Journal of Biochemistry*, 269, 5642-5648.

[12] Tomaselli S., Esposito V., Vangone P., van Nuland N.A, Bonvin A.M., Guerrini R., Tancredi T., Temussi P.A, Picone D., 2006. The Alpha-to-beta Conformational Transition of Alzheimer's A β -(1-42) Peptide in Aqueous Media is Reversible: A Step by Step Conformational Analysis Suggests the Location of Beta Conformation Seeding, *ChemBioChem*, 7, 257-267.

[13] Kim S, Chen J, Cheng T, Gindulyte A, He J, He S, Li Q, Shoemaker BA, Thiessen PA, Yu B, Zaslavsky L, Zhang J, Bolton EE. PubChem 2019 update: improved access to chemical data. *Nucleic Acids Res*, 47(D1):D1102-1109.

[14] Dassault Systèmes BIOVIA, Discovery Studio 2017 R2, Release 2017, San Diego: Dassault Systèmes

- [15] Hanwell, M. D., Curtis, D. E., Lonie, D. C., Vandermeersch, T., Zurek, K. & Hutchison, G.R. 2012. Avogadro: an advanced semantic chemical editor, visualization, and analysis platform. *Journal of Cheminformatics*, 4(17).
- [16] Trott, O. and Olson, A.J. 2010. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J. Comput. Chem.*, 31,455-461.
- [17] Haass C, Selkoe D.J. 2007. Soluble protein oligomers in neurodegeneration: Lessons from the Alzheimer's amyloid beta-peptide. *Nat Rev Mol Cell Biol* 8: 101-112.
- [18] G. M. Morris et al., 2009. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility, *J. Comput. Chem.*, 30:16, 2785-2791.
- [19] Wallace A. C., Laskowski R. A., Thornton J. M. 1995. LIGPLOT: a program to generate schematic diagrams of protein-ligand interactions. *Protein Eng.* 8, 127–134. 10.1093/protein/8.2.127.
- [20] Laskowski R.A., Swindells M.B. 2011. LigPlot?: multiple ligand-protein interaction diagrams for drug discovery. *J Chem Inf Model* 51:2778–2786
- [21] Wallace A.C., Laskowski R.A., Thornton J.M., 1995. LIGPLOT: a program to generate schematic diagrams of protein-ligand interactions. *Protein Eng* 8:127–134
- [22] Salentin, S., Schreiber, S., Joachim Haupt V., Adasme M.F and Michael Schroeder M., 2015. PLIP: fully automated protein-ligand interaction profiler. *Nucleic Acids Research Advance*.
- [23] Kim R., and Skolnick. J., 2008. Assessment of Programs for Ligand Binding Affinity Prediction. *J. Comput. Chem.* 29(8): 1316-1331.