

# Evaluation Of Hygienic Characteristics Of Broiler Carcasses In Two Slaughterhouses And At The Retails Shops

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**Abstract—** The object of this study is to evaluate the microbiological quality of broiler carcasses at the slaughterhouses after the cooling process and in the retail shops in Tirana district. 96 broiler carcasses were collected from two slaughterhouses and from the retail shops in Tirana. For each operator were collected 48 samples, 24 in the slaughterhouse immediately after the cooling process and 24 carcasses from the same operators at the retail shops in Tirana. All samples were collected between March 2015-February 2016, and were tested for mesophil aerobic bacterial counts, E coli counts, and Staph. aureus counts. There were ISO standard methods used for all the analysis performed. Mesophilic bacterial counts for all the samples varied from  $10^2$  cfu/g to  $2.1 \times 10^5$  cfu/g. There were 97.92% in norm and 2.08% of the samples above the acceptance limits of the Albanian regulation. E.coli loads for the 96 samples were between  $<10$  cfu/g and  $1.1 \times 10^6$  cfu/g, with 2.08% of the samples above the acceptance limit for E.coli loads. All the samples were also tested for the St.aureus loads and 34.37% of the samples resulted positive but within the regulation limits.

**Keywords—** broiler; microbiological quality; E.coli; NPA; Staph.Aureus;

## I. INTRODUCTION

Poultry meat is a major source of food borne diseases. According to the World Health organization food borne diseases count for 502 634 cases a year with 30% of this cases caused by poultry meat. Live animals are host to a large number of different microorganisms residing on their skin, feather or alimentary tract. Contamination is possible at any stage of the production process, feather plucking, evisceration, washing of the carcasses, cooling and freezing. Other contamination might be possible from the environment, equipment, operators etc (Mead, 1989). Hygienic characteristics of the poultry carcasses depend on the rearing conditions, health status of the flock, processing conditions, storage conditions at the slaughterhouse, packaging, transportation, and handling of the products at the retail shops and houses before consumption. An efficacious way of preventing food-borne human

diseases is to monitor the microbiological quality of poultry meat and meat products during production, storage and distribution. E According to FRIES (2002), the micro-flora of poultry is transferred from the primary production sites to production lines, and further, by subsequent contamination. Micro flora of crude chicken meat is heterogeneous and originates from slaughtering premises, operators' hands, equipment and outfit, and water and air (ANONYM., 1996). Ensuring safe food supply has been one of the major challenges and concerns for producers, consumers and public health officials in both developing and developed countries. This is because foods excessively contaminated with pathogenic and spoilage micro-organism are undesirable and can cause food borne illnesses. The main capacities for production and processing of poultry meat in Albania are mainly located in the territory of the district of Tirana. These operators are both leading suppliers and distributors of this market area. The study was conducted in the period March 2015-February 2016, in Tirana region. A total of 96 samples were tested, of which 48 samples from operator A and 48 samples from operator B. Samples were taken at the slaughterhouse after the cooling process as well as at the retail markets places supplied from the same operators. Mesophil aerobic count are an indicator of hygienic characteristics of poultry carcasses and E. coli si an indicator of fecal contamination. St. aureus is a common contaminant of poultry meat coming from different origins, with an emphasis of the human strains, which pose the highest risk to the public health (Capita et al, 2002).

## II. MATERIALS AND METHODS

Ninety-six broiler carcasses were collected from two slaughterhouses (24 samples each) and from the retail shops in Tirana (24 samples from each operator). The samples were aseptically collected, and each carcass was placed in a separate, sterile plastic bag. The samples were brought under refrigeration to the laboratory and analyzed within the following 4 h.

### A. Total Mesophilic aerobic counts

The samples were subjected to microbiological analysis according to standard procedures ISO (4833:2003). 25 grams of meat from different parts of each whole carcass was removed with the help of a sterile scalpel and minced manually. It was then thoroughly mixed in 225 ml of Buffered peptone water.

Decimal dilutions were poured into a Petri dish. Total mesophil aerobic count test (TAC) was carried out on Standard plate count agar (PCA media) by pour plate method (in duplicate) and plates were incubated for 48 hours at 37°C. The enumeration of Plates with approximately 25-250 colonies were selected for counting of results. Total numbers of colonies were counted after 48 hours from each plate.

### B. *St.Aureus*

25 g of test sample (meat) was weighted and blended in stomacher for 2 minutes and a gram of the sample was weighed out and homogenized in 225mls buffered peptone water. The samples were diluted at dilution rate 1:10 and 0.1 ml from the diluted sample was inoculated into CTNA (coagulase thermo nuclease agar) and incubated for 48 hours in 37°C+1°C. The colonies of *St aureus* were identified and counted.

## III. RESULTS AND DISCUSSION

### A. Mesophilic Aerobic Counts

Table1. indicates the results for the mesophilic aerobic counts for operator A at the slaughterhouse and at the retail shops. The total mesophilic aerobic counts resulted within the regulation limit for 47 of the samples and over the regulation limit in 1 case at the retail shop.

It is evident that the total mesophilic aerobic counts tend to increase from the slaughterhouse to the retail shop, this is to be expected because of the storage in the retail shops. Samples collected from the Operator A slaughterhouse resulted 18 satisfactory, 6 acceptable with no samples over the acceptable limits. Samples from the same operator collected at the retail shops resulted 7 satisfactory, 16 acceptable, 1 unacceptable.

TABLE I. TOTAL NUMBER OF AEROBIC (NPM) CFU / G

Operatori A		
Month	slaughterh	Market
March 2015	6x10 <sup>3</sup>	1.7x10 <sup>4</sup>
	5x10 <sup>3</sup>	2.5x10 <sup>4</sup>
April	5.5x10 <sup>3</sup>	5.9x10 <sup>3</sup>
	4x10 <sup>3</sup>	2.1x10 <sup>5</sup>
May	6x10 <sup>3</sup>	3x10 <sup>4</sup>
	10 <sup>3</sup>	3.5x10 <sup>3</sup>
June	2.1x10 <sup>3</sup>	4.2x10 <sup>4</sup>
	5.5x10 <sup>3</sup>	2x10 <sup>3</sup>
July	2x10 <sup>3</sup>	2.3x10 <sup>6</sup>
	10 <sup>3</sup>	2.4x10 <sup>3</sup>
August	2.5x10 <sup>3</sup>	5.4x10 <sup>5</sup>
	2.7x10 <sup>3</sup>	2.5x10 <sup>3</sup>
September	1.7x10 <sup>4</sup>	5.2x10 <sup>2</sup>
	1.5x10 <sup>4</sup>	9.1x10 <sup>3</sup>
October	4x10 <sup>3</sup>	1.8x10 <sup>3</sup>
	1.1x10 <sup>3</sup>	3x10 <sup>3</sup>
November	1.5x10 <sup>3</sup>	10 <sup>3</sup>
	1.9x10 <sup>3</sup>	1.7x10 <sup>4</sup>
December	1.7x10 <sup>4</sup>	2.7x10 <sup>3</sup>
	1.5x10 <sup>4</sup>	2.5x10 <sup>3</sup>
January 2016	10 <sup>3</sup>	4x10 <sup>5</sup>
	1.3x10 <sup>3</sup>	9.2x10 <sup>2</sup>
February 2016	4.8x10 <sup>5</sup>	9.9x10 <sup>2</sup>
	1.6x10 <sup>4</sup>	1.1x10 <sup>3</sup>

Table 2 indicates the total mesophilic aerobic counts for the 48 samples collected from the slaughterhouse of the operator B and the retail shops. There 47 samples within the regulation limits and 1 sample collected at the slaughterhouse resulted unacceptable.

TABLE II. TOTAL NUMBER OF AEROBIC (NPM) CFU / G.

Operatori B		
Month	Slaughterh	Market
March 2015	10 <sup>3</sup>	6x10 <sup>2</sup>
	3x10 <sup>4</sup>	3.2x10 <sup>4</sup>
April	1.7x10 <sup>3</sup>	3x10 <sup>3</sup>
	2x10 <sup>4</sup>	4.5x10 <sup>4</sup>
May	8.2x10 <sup>2</sup>	1.8x10 <sup>4</sup>
	4.4x10 <sup>4</sup>	2x10 <sup>4</sup>
June	3x10 <sup>2</sup>	5.4x10 <sup>4</sup>
	4.7x10 <sup>3</sup>	2.3x10 <sup>3</sup>
July	5.66x10 <sup>6</sup>	1x10 <sup>3</sup>
	2.3x10 <sup>3</sup>	1.1x10 <sup>3</sup>
Augusto	3.6x10 <sup>3</sup>	1.1x10 <sup>4</sup>
	5.4x10 <sup>3</sup>	3.5x10 <sup>4</sup>
September	2.8x10 <sup>3</sup>	2.4x10 <sup>4</sup>
	1.5x10 <sup>4</sup>	2.8x10 <sup>4</sup>
October	1.1x10 <sup>4</sup>	2.3x10 <sup>4</sup>
	1.4x10 <sup>4</sup>	2.2x10 <sup>3</sup>
November	10 <sup>3</sup>	2.6x10 <sup>3</sup>
	3.9x10 <sup>3</sup>	1.5x10 <sup>4</sup>
December	2.5x10 <sup>3</sup>	1.8x10 <sup>4</sup>
	2.4x10 <sup>3</sup>	2.2x10 <sup>3</sup>
January 2016	1.7x10 <sup>3</sup>	1.6x10 <sup>3</sup>
	5.5x10 <sup>5</sup>	5.4x10 <sup>5</sup>
February 2016	1.9x10 <sup>3</sup>	1.1x10 <sup>3</sup>
	10 <sup>3</sup>	9.1x10 <sup>2</sup>

In total there were 2.083% (2/96) of the samples unacceptable according to the total mesophilic aerobic counts. It has to be added that the samples were collected randomly at the slaughterhouse and at the retail shops, meaning that the samples at the retail shops could have been stored for longer than 1 day prior to collection.

### B. *E.coli* counts

Forty eight samples were tested for *E.coli* counts from Operator A. Twenty four samples were collected at the slaughterhouse and 24 at the retail shops. Table 3 indicates the *E coil* counts for all the samples tested at the slaughterhouse and at the retail shops. There is to be noticed that the *E coli* counts have a sharp increase in the retail shops because of the extended time of storage and also the extra contamination derived from the environment handling and storing. There is only on case collected from the retail shops which is over the meat regulation limits and all the other samples are within the regulation limits.

TABLE III. ESCHERICHIA COLI COUNTS

Operator A	Slaughterhouse cfu/g	Market cfu/g
March	<10	<10
April	<10	1.5x10 <sup>2</sup>
May	<10	10 <sup>3</sup>
June	4x10	<10
July	1.1x10 <sup>3</sup>	1.1x10 <sup>6</sup>
Augusto	<10	1.1x10 <sup>3</sup>
September	20	9x10 <sup>2</sup>
October	<10	<10
November	20	<10
December	<10	<10
January 2016	<10	2.2x10 <sup>2</sup>
February 2016	<10	3.6x10 <sup>2</sup>
	2.3x10 <sup>2</sup>	5.2x10 <sup>2</sup>

Table 4 indicates the E.coli counts for each samples collected at the slaughterhouse of the operator b and at the retail shops. All the samples resulted within the regulation limits for E.coli counts except on sample from the slaughterhouse, which resulted over the regulation limit.

TABLE IV. ESCHERICHIA COLI COUNTS

Operatori B	Slaughterhouse cfu/g	Market cfu/g
March	<10	<10
April	10 <sup>3</sup>	1.3x10 <sup>3</sup>
May	9.1x10 <sup>2</sup>	1.2x10 <sup>3</sup>
June	<10	<10
July	<10	<10
Augusto	<10	<10
September	1.5x10 <sup>6</sup>	<10
October	<10	<10
November	<10	<10
December	<10	<10
January 2016	<10	8x10 <sup>2</sup>
February 2016	10 <sup>3</sup>	10 <sup>3</sup>
	<10	<10

Table. 5 and 5.1 indicates the St. aureus counts in all samples tested from the operator A and operator B respectively. The St. aureus counts were within the limits in all the 96 samples. The positivity for St. aureus resulted 33/96 (34.37%) of the total number of analyzed samples. It is to be noticed that there is a higher positivity for the samples collected at the retail shops from both operators. This was expected because the carcasses are not packed individually.

with sealed packaging, allowing a higher contamination rate for the samples at the retail shops.

TABLE V. RESULTS OF ST. AUREUS

Operatori A	Slaughterhouse cfu/g	Market cfu/g
March	0	0
April	0	16
May	0	10 <sup>2</sup>
June	0	20
July	70	40
Augusto	2x10 <sup>2</sup>	9x10 <sup>2</sup>
September	20	0
October	10	0
November	0	0
December	0	0
January	0	70
February	0	20
	30	20
	57	0
	0	30
	0	1100
	0	150
	0	0
	0	50

TABLE VI. RESULTS OF ST. AUREUS

Operatori B	Slaughterhouse cfu/g	Market cfu/g
March	0	20
April	0	0
May	0	16
June	0	10
July	0	0
Augusto	0	9x10 <sup>2</sup>
September	0	50
October	9x10 <sup>2</sup>	0
November	50	0
December	0	0
January 2016	0	0
February 2016	0	0
	0	50
	0	0
	0	0
	0	0
	0	0
	0	50
	16	0
	20	0
	0	20
	0	70
	20	0
	0	40
	0	0
	0	0

C. CONCLUSION

Aerobic Mesophilic bacterial counts were within the regulation limit in 97,91% of the samples analyzed and 2.083% of the samples resulted unacceptable.

E coli loads were within limits in 94 samples and unacceptable in 2 samples. There is a slight increase of E.coli counts in the samples collected from the retail shops because of longer storage time and missing of individual packaging.

St aureus counts were within the limits in all samples although there is an increase in positivity and bacterial counts in the samples collected from the retail shops.

Referring to the results obtained, it can be concluded that the hygienic conditions of poultry processed and sold in Albania are within the regulation limits except a small number of cases which resulted unacceptable because of high mesophilic bacterial counts and E. coli counts.

It is also to be noticed that the poultry carcasses should be individually packed with sealed packaging materials in order to avoid further contamination during the storage and distribution at the retail shops.

#### References

- [1] CDC, (Centers for Disease Control and Prevention), 2011. Investigation of a multistate outbreak of human Salmonella I 4, 5, 12 from November 1, 2010, through February 9, 2011.
- [2] Clouser, C.S., Doors, S., Mast, M.G., Knable, S.J. 1995. The role of defeathering in the contamination of turkey skin by Salmonella species and Listeria monocytogenes. J. Poultry Sci., (74):723-731.
- [3] Cruickshank, R., Duguid, J.P., Marmion, B.P., Swain, R.H.A. 1975. Medical Microbiology, 12th ed., Churchill Livingstone Edinburgh, London and New York.
- [4] Delicate, E.R., de Brito, B.G., Gaziri, L.C., Vidotto, M.C., 2003. Virulence-associated genes in Escherichia coli isolates from poultry with Colibacillosis. J. Vet. Microbiol., (94): 97-103.
- [5] Edwards, P.R., Ewing, W.H. 1972. Identification of Enterobacteriaceae, 3rd ed., Burgess Publishing Co., Minneapolis, 85-95
- [6] Eissa, M. Wafaa, 1995. Enterobacteriaceae at various stages of poultry processing. M.V.Sc., Thesis, (Meat Hygiene), Fac. Vet. Med., Alex. Univ
- [7] Mohamed, M. Ghada., Ebraheem, M.L., Thabt, H.M. 2010. Electrophoretic analysis and Immunological characterization of Staphylococcus aureus isolated from chicken meat. Assiut Vet. Med. J., 56: 127-136
- [8] ISO 6887-1: 1999 Microbiology of food and animal feeding stuffs-Preparation of test samples, initial suspension and decimal dilutions for microbiological examination-Part 1: General rules for the preparation of test samples, initial suspension and decimal dilutions
- [9] The method of regulated - ISO / TS 16649-2 Microbiology of food and feed - Horizontal method for the enumeration of Escherichia coli-positive  $\beta$ -technical -glucuronida counting colonies on the plate at 44C using 5-bromo-4-chloro-3-indolyl- $\beta$ -glucuronidase.
- [10] The method of regulated ISO 4833: 2003 - Microbiology of food and feed. Horizontal method for the enumeration of microorganisms - technique of counting colonies on the plate at 30 °C regulated method EN ISO 6579: 2002 / Cor.1: 2004 (E) - Microbiology of food and feed
- [11] Horizontal method for the detection of Salmonella spp Commission regulation EC no. 2073/2005. 2011 on microbiological Criteria for food
- [12] Cardenas et al, 2008, Zahra et al, 2010 Mathematical modeling of microbial growth in ground beef from Argentina. Effect addition of Lactic acid, temperature and packing film.
- [13] [ISO 6887-2: 2003 Preparation of test samples, initial suspension and decimal dilutions for microbiological examination. Part 2: Specific rules for the preparation of meat and meat product
- [14] BARNES, E.M. (1976). Microbiological problems of poultry at refrigerator temperatures - a review. Journal of the Science of Food and Agriculture 27, 777-82.
- [15] Bailey, J. S., J. E. Thomson, and N. A. Cox. 1987. Contamination of poultry during processing. Pages 193–211 in The Microbiology of Poultry Meat Products. F. E. Cunningham, and N.A. Cox ed. Academic Press, Orlando, FL Bilgili, S. G. 1988.
- [16] Effect of feed and water withdrawal on shear strength of broiler gastrointestinal tract. Poult. Sci. 67:845–847 Izat, A. L., M. Colberg, C. D. Driggers, and R. A. Thomas. 1989
- [17] Effect of sampling method and feed withdrawal period on recovery of microorganisms from poultry carcasses. J. Food Prot. 52:480–483 Notermans, S. F., and E. H. Kampelmacher. 1974. Attachment of some bacterial strains to the skin of broiler chickens. Br. Poult. Sci. 15:573–585
- [18] Renwick, S. A., W. B. McNab, H. R. Lowman, and R. C. Clarke. 1993. Variability and determinants of carcass bacterial load at a poultry abattoir. J. Food Prot. 56:694–699.
- [19] McNab, W. B., S. A. Renwick, H. R. Lowman, and R. C. Clarke. 1993. Variability of broiler carcass bacterial load at three abattoirs, as measured by a hydrophobic grid membrane filter interpreter. J. Food Prot. 56:700–705.
- [20] Unnevehr, L. J., and H. H. Jensen. 1996. HACCP as a regulatory innovation to improve food safety in meat industry. Am. J. Agric. Econ. 20:186–201 McNamara, A. M. 1997.
- [21] Generic HACCP applications in broiler slaughter and processing. National Advisory Committee on Microbiological Criteria for Foods. J. Food Prot. 60:579–604
- [22] Thomas, C. J., A. T. McMeekin, and T. J. Patterson. 1987. Prevention of microbial contamination in the processing plant. Pages 163–179 in Elimination of Pathogenic Organisms from Meat and Poultry. F. J. M. Smulders, ed. Elsevier, Amsterdam.
- [23] McMeekin, T. A., and C. T. Thomas. 1978. Retention of bacteria on chicken skin after immersion in bacterial suspension. J. Appl. Bacteriol. 46:383–387
- [24] MEAD, G. C., W. R. HUDSON, M. H. HINTON (1993): Microbiological survey of five poultry processing plants in the UK. Brit. Poultry Sci. 34, 497-503.
- [25] MEAD, G. C. (1989): Hygiene Problems and Control of Process Contamination. In: Processing of poultry. (Mead G. C., Ed.), Elsevier Science Publishers Ltd. 1989, pp. 183-220