# Biosorption Of Chromium (VI) By Different Natural Biomasses

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Abstract—Cr (VI) is a toxic metal, which belongs to the list of priority pollutants due to its mutagenic and carcinogenic properties, defined by the US EPA. Contamination with Cr (VI) comes from electroplating, leather tanning, textile dyeing and metal finishing industries. Recently, a variety of low cost materials has been studied for their ability to remove this metal from aqueous solution with promising results. We studied the Cr (VI) removal capacity in aqueous solution by different natural biomasses, using the diphenylcarbazide method to evaluate the metal concentration in solution and obtaining the highest biosorption of the metal (50 mg/L). It was found that a pH for 1.0, temperature of 60°C, and 5 g of natural biomass, were the optimum conditions of the experiment. In addition, the natural biomasses showed good removal capacity of 1.0 g/L of the metal, at 28°C 60°C, and removed efficiently the metal of earth and water contaminated naturally.

# Keywords—Chromium (VI), Removal, Natural Biomasses, Detoxification

### I. INTRODUCTION

Chromium is regarded as an environmental pollutant due to its wide use in various industrial activities, such as electrolytic plating, leather tanning, explosives manufacturing etc. The stable forms of chromium in the environment are trivalent (Cr (III)) and hexavalent

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chromium (Cr (VI)). Further, Cr (VI) is highly soluble, making it mobile in soil and aquatic environments, with consequent toxicity ecosystems. Chromium in their different forms can be use in the production of steel alloys and other metals chromed, for dyes and pigments, and the preservation of leather and wood. It can also be find naturally in the soil. The primary forms of chromium found in nature are chromium (III) and chromium (VI) and these forms are converted to each other depending on environmental conditions [2]. Cr (VI) is consider the most toxic form of chromium, and is usually associated with oxygen as chromates  $(CrO_4^{-2})$  and dichromates  $(Cr_2O_7^{-2})$  [3], which due to its high solubility are highly mobile in soil environments and water [4]. Moreover, Cr (III) is in the form of oxides, hydroxides or poorly soluble sulfates, by which it is much less mobile, and there joined organic matter in the soil and aquatic environments [5, 6]. Cr (VI) is a strong oxidizing agent, and in the presence of organic matter is reduced to Cr (III); this transformation is faster in acidic environments [3]. However, high levels of Cr (VI) may exceed the reducing capacity of the environment and thus can persist as a contaminant. It has been established now that various chromium compounds as oxides, chromates and dichromates, are environmental contaminants in water, soil, and industrial effluents, because this metal is widely used in various manufacturing, such as electrolytic plating, explosives manufacturing, leather tanning, metal alloy, dyes and pigments manufacturing, etc. [1, 5].

There are studies of many methods for removal of chromium ion present in water industrial waste, for example: ion exchange on resins, coagulationflocculation, adsorption on activated carbon, reduction, chemical precipitation, sedimentation, etc., [7], which in most cases are expensive or inefficient, especially when the concentration of these ions is very low [8]. Therefore arise emerging technologies such as biosorption, the process of attracting various chemical species by biomass (live or dead), by physicochemical mechanisms as adsorption or ion exchange [9]. Recently, varieties of low cost materials have been studied for their ability to remove Cr (VI) from aqueous solution and promising results are shown. Among these low cost adsorbents are dead microorganisms, clay minerals, agricultural wastes, industrial wastes and various other low cost materials [1, 10]. Thus, there is a need to develop or find innovative low cost adsorbents with an affinity towards metal ions for the removal of Cr (VI) from aqueous solution, which leads to high adsorption capacity [11]. The objective of this study was to analyze in vitro biosorption of Cr (VI) by different natural biomasses.

II. EXPERIMENTAL

### A. Biosorbent used

The Mangifera indica, Musa paradisiaca, Citrus paradise, Cucumis melo, Cucurbita máxima shells, and the sawdust of Pinus sylvestris, were obtained from the fruits harvested and offered in the market place Republic, and of different carpentry, respectively, between the months of June to September in 2014, of the capital city of San Luis Potosí, S.L.P. México. To obtain the biomasses, the shells and sawdust were washed with trideionized water 72 hours under constant stirring, with water changes every 12 hours.

Subsequently, boiled 1 hour to remove traces of the fruit and dust, and were dry at 80°C for 12 hours in the oven, ground in blender and stored in amber vials until use.

### B. Biosorption studies and determination of hexavalent, trivalent, and total Cr

In these studies, was used 1 g of dried biomass mixed with 100 mL, containing 50 mg/L of the metal bearing solution in an Erlenmeyer flask at the desired temperature and pH. The flasks were agitated on a shaking bath Yamato BT-25 model at different times. Samples of 5 mL were taken at different times and centrifuged at 3000 rpm for 5 min.

The supernatant liquid was separated and analyzed for Cr (VI) ions. Hexavalent Chromium and trivalent Chromium were quantify by a spectrophotometric method employing diphenylcarbazide and chromazurol S, respectively [12, 13], total Chromium was determine by electrothermal atomic absorption spectroscopy [12]. The information shown in the results section are the mean from three experiments carried out by triplicate.

### C Statistical analysis

The statistical aspect considered the removal times using the Microsoft®Excel program and EPI INFO 7.

The OR of each group compared with a statistical significance of p < 0.05 was calculated.

### III. RESULTS AND DISCUSSION

### A. Effect of incubation time and pH

The optimum time and pH for Cr (VI) removal for the biomasses of M. indica, M. paradisiaca, C. paradise, C. melo, C. máxima shells, and the sawdust were 1.16, 60, 120, 180, 480, and 960 minutes, resepecively, and pH 1.0, at constant values of biosorbent dosage (1 g/100 mL), with an initial metal concentration (50 mg/L), and temperature of 28°C (Figure 1). It was used a pH meter Corning Pinnacle 530 model and we use nitric acid 1M to maintain the pH. M. indica biomass removed 25 times faster chromium VI compared with M. paradise biomass, 9 times faster than the biomass of M. indica, 50 times faster than biomass of C. melo (p < 0.001), 4 times faster than the C. máxima biomass (p < 0.05) and 33.33 times faster than sawdust biomass. (p < 0.001). In the literature [14], report an optimum time of 60 min for the removal of lead by orange shell, 30 min and 2 hours for the removal of Cr (VI) by the Citrus reticulata shell and eucalyptus bark [15, 16]. Changes in the permeability of unknown origin, could partly explain the differences founded in the incubation time, providing greater or lesser exposure of the functional groups of the cell wall of the biomass analyzed [1, 10].

Adsorption efficiency of Cr (VI) was observe a maximum at pH 1.0 with all the biomasses analyzed, and this is like to most reports [1, 10, 15, 17, and 18] This was due to the dominant species ( $\text{CrO}_4^{2^-}$  and  $\text{Cr}_2\text{O}_7^{2^-}$ ) of Cr ions in solution, which were expect to interact more strongly with the ligands carrying positive charges [19].

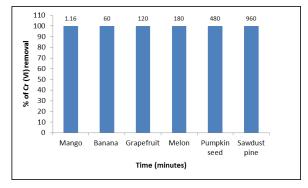


Figure 1.100% removal of Chromium (VI) (50 mg/L), pH 1.0, different natural biomasses, 28°C, 1 g biomass, 100 rpm.

### B. Effect of the temperature

Temperature was found to be a critical parameter in the bioadsorption of Cr (VI) (Figure 2). To maintain constant the temperature in all experiments, we use a shaking bath Yamato BT-25 model. The total removal was observed at 28°C and 60°C, and the most efficient biomass was *M. indica* at 1.16 and 5 min, respectively of incubation time. *C. máxima* biomass removal 15 times faster the concentration of chromium VI that *M. paradisiaca* biomass (P <0.05) and 7.5 times faster than grapefruit (P <0.05). No statistically significant difference between the biomasses of *M. indica*, sawdust and *C. maxima*. Both, the *C. maxima* and sawdust biomasses increase the speed of removal at higher concentrations of chromium VI; however, there is no statistically significant difference between them.

This results are coincident for tamarind shell with 95% of removal at 58°C and 3 hours [20], for the adsorption of cadmium (II) from aqueous solution on natural and oxidized corncob (40°C and 5 days) [21], but these are different for the mandarin waste [22], Caladium bicolor (wild cocoyam) biomass [23], and *Saccharomyces cerevisiae* [24]. The increase in temperature increases the rate of removal of Chromium (VI) and decrease the contact time required for complete removal of the metal, to increase the redox reaction rate [20].

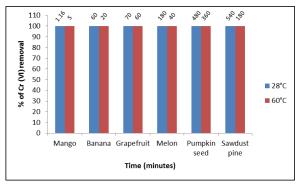


Figure 2. 100% removal of Chromium (VI) (50 mg/L), pH 1.0, different natural biomasses, 1 g biomass, 100 rpm,  $28^{\circ}$ C and  $60^{\circ}$ C.

### C. Effect of initial metal concentration

At low metal concentrations (200 mg/L), we observe the best results for removal, with all the biomasses analyzed, at 60°C and 28°C, (200 mg/L and 1000 mg/L) respectively (Figures 3a, and 3b). Between biomasses of *C. paradisiaca*, *C. paradisi*, and sawdust, we do not found difference in removal at a concentration of 200 mg/L of chromium VI. Sawdust and *C. maxima* biomasses removal five times faster 1000 mg/L of chromium VI concentration what *C. paradisi* biomass (p <0.05), whereas no significant difference between Sawdust, *C. maxima*, and *C. paradisiaca*.

In addition, we observe the development of a bluegreen and white precipitate, which changes more rapidly at higher temperatures (date not shown). The results are coincident for *C. reticulata* shell [15, 20]. With respect to other biomasses, most authors report lower removal efficiencies of metal, for example: 45 mg/L for eucalyptus bark [16], 13.4 and 17.2 mg/L for bagasse and sugar cane pulp, 29 mg/L coconut fibers, 8.66 mg/L for wool [25], 25 and 250 mg/L of chitin and chitosan [26], and 1 mg/L for cellulose acetate [27]. The increase in initial concentration of Cr (VI), results in the increased uptake capacity and decreased in the percentage of removal of the metal. This was due to the increase in the number of ions competing for the available functional groups on the surface of biomass [20].

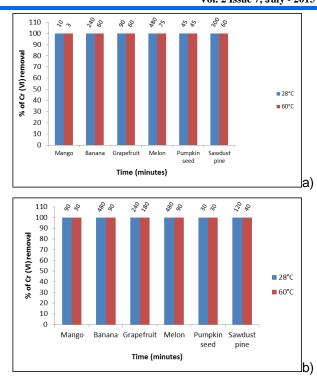


Figure 3. 100% removal of Chromium (VI) (28°C and 60°C), different natural biomasses, 1 g of natural biomass, pH 1.0, 100 rpm, a) 200 mg/L, b) 1000 mg/L.

### D. Effect of biosorbent dose

The influence of biomass on the removal capacity of Cr (VI) is depict in Figure 4. If we increase, the amount of biomass also increases the removal of the metal in solution, with more biosorption sites of the same, because the amount of added biosorbent determines the number of binding sites available for metal biosorption [28]. With 5 g, M. indica biomass removed 30 times faster than biomass of C. reticulata (p <0.001), 66.66 times faster than biomass of C. maxima (p < 0.001), and 125 times faster sawdust biomass (p <0.001). Similar results have been reported for modified corn stalks [31], C. reticulata shell [15], and Mucor hiemalis and Rhizopus nigricans, although latter with 10 g of biomass [29, 30], but they are different from those reported for wastes biomass of mandarin (gabasse), with an optimal concentration of biomass of 100 mg/L [22].

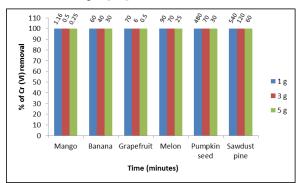


Figure 4. 100% removal of Chromium (VI) (50 mg/L), with 1,3 and 5 g of different natural biomasses, 28°C, pH 1.0, 100 rpm.

# E. Time course of Cr (VI) decrease and Cr (III) production by different solutions

The ability of the different biomasses of to lower the initial Cr (VI) of 1.0 g/L and Cr (III) production in solution was analyzed. Figure 5 shows that the biomass exhibited a remarkable efficiency to diminish Cr (VI) level with the concomitant production of Cr (III) in the solution (indicated by the formation of a bluegreen color and a white precipitate, and his determination for Cromazurol S, date not shown) [13]. Thus, between 35 and 720 min of incubation, the biomasses analyzed, caused a drop in Cr (VI) from its initial concentration of 1.0 g/L to almost undetectable levels, and the decrease level occurred without change significant in total Chromium content. As expected, total Chromium concentration remained constant over time, in solution control. These observations indicate that these biomasses are able to reduce Cr (VI) to Cr (III) in solution. Furthermore, as some of the biomasses contains vitamin C and some carbohydrates, we found that vitamin C and cystine reduce faster Cr (VI) to Cr (III), and could be very important part in the metal reduction, confirming some reports in the literature [3, 15, 32, 33, and 34]. There are two mechanisms by which chromate could be reduced to a lower toxic metal. Oxidation state by an enzymatic reaction. Currently, we do not know whether the shell biomass used in this express study and Cr (VI) reducing enzyme(s). Further studies are necessary to extend our understanding of the effects of coexisting ions on the Cr (VI) reducing activity of the biomass reported in this study. In addition, Cr (VI) reducing capability has been described in some reports in the literature [3, 7, 8, 32, 35-38].Biosorption is the second mechanism by which the chromate concentration could be reduced, because the biomass shell can be regarded as a mosaic of different groups that could form coordination complexes with metals, and our observations are like to the most of the reports in the literature [3, 7, 8, 32, 35-38].

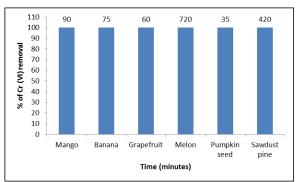


Figure 5. Time-course of Cr (VI) decrease and Cr (III) production in solution with 1.0 g/L Cr (VI). 28°C, pH 1.0, 100 rpm.

### F. Removal of Cr (VI) in industrial wastes with biomass of C. melo shell

We adapted a water-phase bioremediation assay to explore possible usefulness of the biomasses for eliminating Cr (VI) from industrial wastes, the biomass was incubate with non-sterilized contaminated soil and water containing 297 mg Cr (VI)/g, and 155 mg Cr(VI)/L, suspended in trideionized water. It was observe that in 7 days of incubation with the biomasses, the Cr (VI) concentration of soil and water samples decrease between 57.2% and 100% (Figure 6), and the decrease level occurred without change significant in total Chromium content during the experiments. In the experiment carried out in the absence of the biomass, the Cr (VI) concentration of the soil samples decreased by about of 18% (date not shown); this might be caused by indigenous microflora and (or) reducing components present in the soil.

The biomasses of C. melo, C. maxima and sawdust, not removal 100% of the metal in 7days. however, biomasses of M. indica, M. paradisiaca and C. reticulata, removal the 100% of the metal, and there is no statistically significant difference. The chromium removal abilities of this biomasses, are equal or better than those of other reported, for example C. reticulata shell [15], M. Americana [17] and Candida maltose RR1 [39]. In particular, these biomasses were superior to the other biomass because they has the capacity for efficient chromium reduction under acidic conditions. Many of the Cr (VI) reduction studies were carry out at neutral pH [40]. Aspergillus niger also has the ability to reduce and adsorb Cr (VI) [40]. When the initial concentration of Cr (VI) was 500 ppm, A. niger mycelium removed 8.9 mg of chromium/g dry weight of mycelium in 7 days.

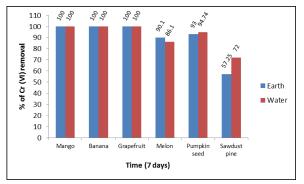


Figure 6. Biorremediation of Earth and Water contaminated with 297 mg Cr (VI)/ g of Earth and 400 mg/L of Water respectively, 5 g of natural biomass. 28°C, 100 rpm.

### IV. CONCLUSIONS

The biomasses analyzed, showed complete capacity of biosorption of 1.0 g/L Cr (VI) in solution at different time of incubation, at  $28^{\circ}$ C, 100 rpm with 1 g of biomass, besides thes removes the metal in situ (7 days of incubation, 5 g of biomass), in soil and water contaminated, respectively. These results suggest the potential applicability of these biomasses for the remediation of Cr (VI) from polluted soils in the fields.

### V. REFERENCES

[1] M. Ahemad, "Bacterial mechanisms for Cr(VI) resistance and reduction: an overview and recent advances". Folia Microbiologica, vol. 59, pp. 321–332. 2014.

[2] United States Environmental Protection Agency.

[3] H. Seng, and Y.T. Wang, "Biological Reduction of Chromium by *E. coli*", J. Environ. Eng., vol. 120(3), pp. 560-572, 1994.

[4] T.L. Marsh, and M.J. McInerney, "Relationship of Hydrogen Bioavailability to Chromate Reduction in Aquifer Sediments", Appl. Environ. Microb., vol. 67(4), pp. 1517-1521, 2001.

[5] J.O. Nriagu, and E. Nieboer, "Chromium in the natural and human environments. Wiley-Interscience, New York; 1988.

[6] G. Lofroth, and B.N. Ames, "Mutagenicity of Inorganic Compounds in *Salmonella typhimurium*: Arsenic, Chromium and Selenium", Mutat. Res., vol. 53(1), pp. 65-66, 1978.

[7] D. Park, Y.S. Yun, H.Y. Cho, and J.M. Park, "Chromium Biosorption by Thermally Treated Biomass of the Brown Seaweed, *Ecklonia* sp", Ind. Eng. Chem. Res., vol. 43(26), pp. 8226-8232, 2004.

[8] Y. Sahin, and A. Öztürk, "Biosorption of Chromium (VI) lons from Aqueous Solution by the Bacterium *Bacillus thuriengensis*", Process Biochem., vol. 40(5), pp. 1895-1901, 2005.

[9] B. Volesky, and Z.R. Holan. "Biosorption of heavy metals". Biotechnology Progress 1995; vol. 11, 235-250. 1995.

[10] C. Tejada-Tovar, Á. Villabona-Ortiz y L. Garcés-Jaraba, "Adsorción de metales pesados en aguas residuales usando materiales de origen biológico". Tecno Lógicas. vol. 18, (34), pp. 109-123. 2015.

[11] R. Kumar, N.R. Bishnoi, and G.K. Bishnoi, "Biosorption of chromium (VI) from aqueous solution and electroplating wastewater using fungal biomass", Chem. Eng. J., vol. 135, pp. 202–208, 2008.

[12] A.E. Greenberg, L.S. Clesceri, and A.D. Eaton, "Standard Methods for the Examination of Water and Wastewater", American Public Health Association, Washington, DC, USA, 18th edition, 1992.

[13] R.P. Pantaler, and I.V. Pulyaeva, "A spectrophotometric study of complexation between chromium and chromazurol S", J. Anal. Chem. (Moscow), vol. 40, pp. 1634-1639, 1985.

[14] A.B. Pérez, V. Meseguer, J.F. Zapata, M. Ortuño, J. Aguilar, S. Sáez, and M. Lloréns, "Removal of cadmium from aqueous solutions by adsorption onto orange waste", J. Hazard. Mater., vol. 139, pp. 122-131, 2007.

[15] I. Acosta, E. Coronado, J.F. Cárdenas, J. Tovar, and V.M. Martínez, "Hexavalent chromium removal by *Citrus reticulata* shell", J. Nat. Sci., vol. 1(1), pp. 29-39, 2013.

[16] V. Sarin, and K.K Pant, "Removal of chromium from industrial waste by using eucalyptus bark", Bioresour. Technol., vol. 97, pp. 15-20, 2006.

[17] I. Acosta, P. Sandoval, D. Bautista, N. Hernández, J.F. Cárdenas, and V.M. Martínez, "Bioadsorción de Cromo (VI) por la cáscara de Mamey (*Mammea americana* L.)", Av. Cienc. Ing., vol. 3(2), pp. 1-9, 2012.

[18] I. Acosta-Rodríguez, R. Martínez-Pérez, J.F. Cárdenas-González, M.G. Moctezuma-Zárate, and

V.M. Martínez-Juárez, Hexavalent Chromium Removal by *Litchi chinensis* Sonn Peel", Amer. J. of Biochem. and Biotech., vol 8 (1), pp. 7-13, 2012.

[19] V.K. Gupta, A.K. Srivastava, and N. Jain, "Biosorption of chromium (VI) from aqueous solutions by green algae *Spirogyra* species", Water Res., vol. 35, pp. 4079–4085, 2001.

[20] G.S. Agarwal, H. Kumar, and S. Chaudari. "Biosorption of aqueous chromium (VI) by *Tamarindus indica* seeds", Bioresour. Technol., vol. 97, pp. 949-956, 2006.

[21] R. Leyva, L.A. Bernal, and I. Acosta, "Adsorption of cadmium (II) from aqueous solution on natural and oxidized corncob", Sep. Purif. Technol., vol. 45, pp. 41–49, 2005.

[22] A. Zubair, H.N. Bhatti, M.A. Hanif. and F. Shafqat, "Kinetic and equilibrium modeling for Cr(III) and Cr(VI) removal from aqueous solutions by *Citrus reticulate* waste biomass", Water, Air Soil Pollut., 191, pp. 305-318, 2008.

[23] M.H. Jnr, and A.I. Spiff, "Effects of temperature on the sorption of Pb2+ and Cd2+ from aqueous solution by Caladium bicolor (wild cocoyam) biomass", Electron. J. Biotechnol., vol. 8(2), pp. 162–169, 2005.

[24] A. Ozer, and D. Ozer, "Comparative study of the biosorption of Pb (II), Ni (II) and Cr (VI) ions onto Saccharomyces cerevisiae: Determination of biosorption heats", J. Hazard. Mater., vol. 100, pp. 219–229, 2003.

[25] M. M. Dakiky, M. Khamis, A. Manassra, and M. Mereb, "Selective adsorption of chromium (VI) in industrial wastewater using low-cost abundantly available adsorbents", Adv. Environ. Res., vol. 6, pp. 533–540, 2002.

[26] Y. Sag, and Y. Aktay, "Kinetic studies on sorption of Cr (VI) and Cu (II) ions by chitin, chitosan and *Rhizopus arrhizus*", Biochem. Eng. J., vol. 12, pp. 143-153, 2002.

[27] G. Arthanareeswaran, P. Thanikaivelan, N. Jaya, D. Mohan, and M. Raajenthiren, "Removal of chromium from aqueous solution using cellulose acetate and sulfonated poly (ether ketone) blend ultrafiltration membranes", Biochem. Eng. J., vol. 12, pp. 43-153, 2002

[28] C. Cervantes, J. Campos, S. Devars, F. Gutiérrez, H. Loza, J.C. Torres, and R. Moreno, "Interactions of chromium with microorganisms and plants", FEMS Microbiol. Rev., vol. 25, pp. 335-347, 2001.

[29] N. Tewari, P. Vasudevan, and B. Guha, "Study on biosorption of Cr(VI) by *Mucor hiemalis*", Biochem. Eng. J., vol. 23, pp. 185-192, 2005.

[30] R.S. Bai, and T.E. Abraham, "Biosorption of Chromium (VI) from Aqueous Solution by *Rhizopus nigricans*", Bioresource Technol., vol. 79 (1), pp. 73-81, 2001.

[31] S. Chen, Q. Yue, B. Gao, Q. Li, and X. Xu, "Removal of Cr(VI) from aqueous solution using modified corn stalks: Characteristic, equilibrium, kinetic and thermodynamic study", Chem. Eng. J., vol. 168, pp. 909- 917, 2011.

[32] W.A. Smith, W.A. Apel, J.N. Petersen, and B.M. Peyton, "Effect of Carbon and Energy Source on Bacterial Chromate Reduction", Biorem. J., vol. 6(3), pp. 205-215, 2002.

[33] X.R. Xu, H.B. Li, J.D. Gu, and X.Y. Li, "Kinetics of the reduction of Chromium (VI) by Vitamin C", Environ. Toxicol. Chem., vol. 24(6), pp. 1310-1314, 2005.

[34] L. Yong, X. Xin, and H. Ping, "Remediation of Cr (VI) in solution using vitamin C", J. Zhejiang Univ. Sci., vol. 6B (6), pp. 540-542, 2005.

[35] D.L. Arévalo, J.F. Cárdenas, V.M. Martínez, and I. Acosta, (2013), "Hexavalent Chromate Reductase Activity in Cell Free Extracts of Penicillium sp", Bioinorg. Chem. Appl. Vol. 2013, Article ID 909412, 6 pages. http://dx.doi.org/10.1155/2013/909412

[36] M.V. Aldrich, J.L. Gardea, J.R. Peralta, and J.G. Parsons, "Uptake and Reduction of Cr(VI) to Cr(III) by Mesquite (Prosopis ssp.): Chromate-Plant Interaction in Hydroponics and Solid Media Studied Using XAS", Environ. Sci. Technol., vol. 37, pp. 1859-1864, 2003.

[37] N. Fiol, C. Escudero, and I. Villaescusa, "Chromium sorption and Cr(VI) reduction to Cr(III) by grape stalks and yohimbe bark", Bioresour. Technol., vol. 99, pp. 5030-5036, 2008.

[38] R. Ramírez, C. Calvo, M. Avila, P. Lappe, M. Ulloa, R. Vázquez, and J.F. Gutiérrez, "Cr(VI) reduction in a Chromate-resistant strain of Candida maltose isolated from the leather industry", Antonie van Leeuwenhoek, vol. 85, pp. 63-68, 2004.

[39] R. Ramírez, C. Calvo, M. Avila, P. Lappe, M. Ulloa, R. Vázquez, and J.F. Gutiérrez, "Cr(VI) reduction in a Chromate-resistant strain of Candida maltose isolated from the leather industry", Antonie van Leeuwenhoek, vol. 85, pp. 63-68, 2004.

[40] T. Fukuda, Y. Ishino, A. Ogawa, K. Tsutsumi, and H. Morita, "Cr(VI) reduction from contaminated soils by *Aspergillus* sp. N2 and *Penicillium* sp. N3 isolated from chromium deposits", J. Gen. Appl. Microbiol., vol. 54, pp. 295-303, 2008.